

Insects associated with exposed decomposing bodies in the Colombian Andean Coffee Region

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ABSTRACT. Insects associated with exposed decomposing bodies in the Colombian Andean Coffee Region. In Colombia, mainly classic forensic medicine methods were used to clarify crimes until 2004. However, other disciplines, including forensic entomology, started to be considered only after the New Accusatory System introduction in Bogotá and the Coffee Region in 2005. In order to provide tools for obtaining evidentiary material elements in judicial trials, it is presented here the succession of insects throughout the decomposition process of an exposed carcass of *Sus scrofa* Linnaeus 1758 (Suidae) and the Occurrence Matrix of colonizing species. This process was evaluated under ambient conditions in the Andean rural area of the city of Pereira, in the Mundo Nuevo district, located in a pre-montane Wet Forest area, from October to November 2006. A sampling period of 27 days and 3198 individuals were collected. We found these colonizing species in the following stages of decomposition: *Lucilia eximia* (Wiedemann, 1819) fresh; *Hemilucilia semidiaphana* (Rondani, 1850), *Oxelytrum discicolle* (Brullé, 1840), and *Cochliomyia macellaria* (Fabricius 1775) bloated; *Chrysomya albiceps* (Wiedemann 1819), *Comptosyriops verena* (Walker, 1849), *Ophyra aenescens* (Wiedemann, 1830) and *Musca domestica* Linnaeus, 1758 active; *Fannia* sp. advanced and *Stearibia nigriceps* (Meigen, 1826) remains. This study provides support tools to define the Post Mortem Interval that may be used by experts from government institutions and laboratories officially accredited.

KEYWORDS. Andean central mountain chain; forensic entomology; Post-Mortem Interval.

RESUMO. Insetos associados com corpos em decomposição na Região Cafeeira Andina Colombiana. Na Colômbia predominou até o ano 2004 a utilização da medicina forense como ferramenta que facilita o processo de esclarecimento de fato criminoso. Contudo, outras disciplinas, entre elas a entomologia forense, começaram a ser utilizadas após o estabelecimento do Novo Sistema Acusatório no ano de 2005 em Bogotá e no Eje Cafetero (região cafeeira). Com o objetivo de gerar novas ferramentas dentro de processos judiciais, é apresentada aqui a sucessão de insetos em carcaça exposta de *Sus scrofa* Linnaeus 1758 (Suidae) e a Matriz de Ocorrência das espécies colonizadoras. O estudo foi realizado em zona rural da cidade de Pereira, localizada em floresta úmida pré-montana, de outubro a novembro de 2006. O período amostral foi de 27 dias e foram coletados 3198 indivíduos, sendo as seguintes as espécies colonizadoras dos estágios de decomposição: *Lucilia eximia* (Wiedemann, 1819) fresco; *Hemilucilia semidiaphana* (Rondani, 1850), *Oxelytrum discicolle* (Brullé, 1840), *Cochliomyia macellaria* (Fabricius 1775) inchado; *Chrysomya albiceps* (Wiedemann 1819), *Comptosyriops verena* (Walker, 1849), *Ophyra aenescens* (Wiedemann, 1830) e *Musca domestica* Linnaeus, 1758 ativo; *Fannia* sp. avançado e *Stearibia nigriceps* (Meigen, 1826) restos. Este estudo gera ferramentas para definição do Intervalo Pós-Morte, que poderá ser utilizado por peritos de instituições do governo e laboratórios oficialmente acreditadas.

PALAVRAS-CHAVE. Cordilheira central andina; entomologia forense; Intervalo Pós-Morte.

Forensic entomology is a very important and frequently used tool for clarifying murders, suicides and other criminal acts. Seventy-two hours after death, this tool becomes the most accurate and sometimes the only tool for determining the time of death (Anderson & VanLaerhoven 1996). Likewise, it may determine whether or not a body has been moved from one place to another, provide information about the place of death and thus, identify both the criminal and the victim (Anderson 2001).

A decomposing body is considered an ephemeral microhabitat that contains a high quality of nutrients. This exists for a short time, is small and is spatially dispersed, where more than one generation of insects develops, which explains the diverse congregation of insects and other invertebrates that colonize it (Hanski & Cambefort 1991). This is how carrion insects use the resource to support their growth, sustenance and reproduction (Schowalter 2000), colonizing the microhabitat for a limited time, and generating an

entomological succession with a regular appearance pattern.

Knowledge of this succession pattern and development rates of each species found provides elements for estimating the Post Mortem Interval (PMI) (Benecke 1998 LaMotte & Wells 2000).

In Colombia, mainly classic forensic medicine methods were used to clarify crimes until 2004. However, other disciplines, including forensic entomology, started to be considered only after the New Accusatory System introduction in Bogotá and the Coffee Region in 2005 (Law 906 of 2004) (Peñuela 2005). Despite being a recently applied tool in this country, entomological succession studies, growth curves and life cycles of forensically-important insect species and entomotoxicology studies have been conducted, all focused on obtaining relevant data for forensic entomology.

With regard to entomological succession, diverse works have been developed in Colombia using white pig carcasses in various altitudes and geographic areas (Wolff *et al.* 2001-

Medellín, 1.500 m.; Pérez *et al.* 2005- Medellín urban area; Martínez *et al.* 2007- Chingaza, Andean Highlands, 3.035 m.; Segura *et al.* 2009- Bogota semi-rural area, 2.700 m.; D. Grisales (unpublished data) - Morrosquillo Gulf, dry tropical forest, 0 m.. With regard to the identification of forensically-important insect species in human corpses, the work by Barreto *et al.* (2002) in the city of Cali, located at 995 m. above sea level. In the area of entomotoxicology, Wolff *et al.* (2004, 2006) conducted studies for quantifying and detecting Parathion and Propoxur, respectively, in forensically-important species.

Studies of life history characteristics of diverse species used in forensic entomology, Camacho & Usaquén (2004) calculated growth curves for the species *Lucilia sericata* (Meigen, 1826) in human liver in the city of Bogotá at 2.600 m. Camacho (2005) identified the entomological succession and growth curves for *Calliphora vicina* Robineau-Desvoidy 1830 in Bogotá. The same was done for the species *Lucilia eximia*, *Cochliomyia macellaria*, *Chrysomya albiceps*, *Chrysomya megacephala* (Fabricius, 1794) and *Calliphora nigribasis* Macquart, 1851, bred in different altitudes by Vélez & Wolff (2008). There are also notes on the behavior and distribution of forensically-important Calliphoridae in Colombia (Wolff & Vélez 2007).

Considering the absence of forensic entomology studies in the Andean Coffee Region, the herein study is considered pioneer since it presents the succession of insects associated with the decomposition process of carcasses and the occurrence matrix (presence-absence) of forensically-important insects in the rural area of the city of Pereira, Risaralda Department. The study provides support tools to define the Post Mortem Interval that may subsequently be used by state institutions, such as the District Attorney's Office and the Medical Examiner's Office.

MATERIAL AND METHODS

The study area is located in the country house *Finca La Acuarela* (altitude of 1,550 m), in the Mundo Nuevo district (4°49'N75°42'W), rural area of the city of Pereira (Risaralda Department), known as part of Colombia's Coffee Belt. It has an average temperature of 21°C, an average annual precipitation of 2,750 mm, with periods of rain concentration in April-May and September-November. The wettest months are April and October (812, 6 mm) and the driest is February (32 mm) (Aguilar & Rangel 1994).

This area belongs to the Sub-Andean region (1,100 m. – 2,500 m), with great antropic activity on vegetable formations and the most inclined areas with relicts of forests (Aguilar & Rangel 1994), belonging to the Pre-Montane Wet Forest life area, according to Holdridge (1987).

A 25-Kg white pig carcass (*Sus scrofa* L.) was used as the decomposition model. The pig was killed with a stab to the heart (Martínez *et al.* 2007) in October 2006. Immediately following the death of the pig, it was placed in a metal cage (approx. 60cm. x 40cm. x 40 cm.) to allow access of insects to the carcass and avoid the attack of carrion-eating vertebrates (Wolff *et al.* 2001). The cage was placed in an

open field and sunlit area. The ecosystem that surrounded the collection site was mainly *Guadua angustifolia* Kunth, 1880 (Bambusaceae), *Heliconia* sp. (Musaceae) crop and a relict of Andean forest.

The samplings were carried out from October 9th to November 3rd 2006, until the bodies were found in remains. Immature (eggs, larvae and pupae) and adult insects were collected during the first two weeks, three times a day (7:00, 12:00 and 17:00). When a remains stage of decomposition was recognized, a single collection was performed at 12:00, until the process ends. In each sampling, adult insects found flying over and/or posing on the body were collected with an entomologic net. Immature insects, other adults feeding on the body and those found under the body and on the ground were collected manually.

During each sampling, body, rectal, and ambient temperatures were recorded with a thermometer. Observable physical changes such as smell, swelling, liquids, gases, etc., were recorded. The carcass's weight was recorded in the morning when the first samples were made.

Adult insects were killed with ethyl acetate in glass jars and were mounted in the laboratory. The immatures were kept in 70% alcohol.

In order to know colonizing flies of the decomposition process, eggs found in the first three days were placed in 200 gr. of raw pork liver, in a plastic container with a screen filter top. Every 5 hours, a sample of 10 larvae was taken to the laboratory until completing the cycle. The samples were killed in 70% alcohol (larvae) and in ethyl acetate (adults).

Both the adult and immature insects were identified with the keys of Borror *et al.* (1989), McAlpine *et al.* (1981, 1987), Dear (1985), White (1983), Greenberg & Szyska (1984), Liu & Greenberg (1989), Queiroz & Carvalho (1987), Wells *et al.* (1999), Stehr (1987), Smith (1986) and Carvalho (2002). For identification of the immature insects, previous clarification and permanent mounting in Canada balsam was done. All of the samples collected were codified and deposited in the Laboratorio de Colecciones Entomológicas, Universidad de Antioquia (CEUA).

Data Analysis

To verify the statistical differences between abundance of individuals collected in each decomposition stage a Wilcoxon Matched Pairs Test (WMP) was performed, with the Statistica "v 7" program. Only data with $p < 0.05$ were considered significant.

With the information about the colonizing and the most abundant species in each stage of decomposition, an occurrence matrix (presence-absence) was done with immature dipterans and both immature and adult coleopterans, which are the most abundant and forensically important groups (Schoenley *et al.* 1992).

RESULTS

3,198 specimens (immature and adult) belonging to 5 orders, 20 families, 30 genera and 36 species were collected

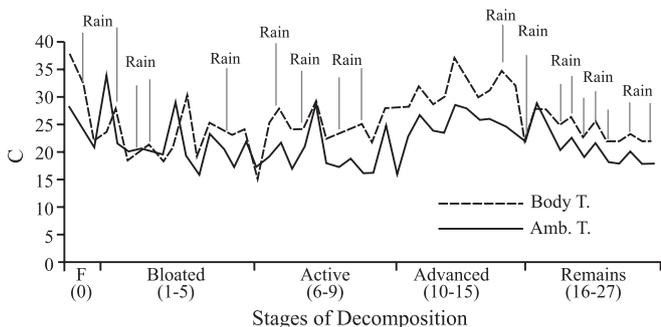


Fig. 1. Daily temperature variations related with stage of decomposition and rainy days.

(Table I). Of that amount, 2,837 individuals (88.7%) were Diptera; 245 (7.66%) Coleoptera; 60 (3.12%) Hymenoptera; 52 (1.6%) Acari; and 4 (0.12%) Hemiptera. Within Diptera, 2 families predominated: Calliphoridae (69.13%) and Muscidae (19.38%). The remaining percentage corresponds to specimens belonging to Syrphidae, Fanniidae and Tephritidae (Table II, III).

Coleoptera was the second most abundant order and 4 families were the most abundant: Desmestidae (3.56%), Silphidae (2.12%), Staphylinidae (1.09%) and Histeridae (0.75%). (Tables II, III).

Stages of decomposition and entomological succession.

Five stages of decomposition were identified (fresh, bloated, active, advanced and remains) considering changes in body temperature, biomass reduction and physical changes in the pig carcass (Wolff *et al.* 2001; Martinez *et al.* 2007).

Abundance of individuals collected in each of the stages of decomposition showed significant differences between them ($p \leq 0.000$). Only the abundance between fresh and remains didn't show significant difference ($p = 0, 434757$).

Fresh (Day 0).

In this stage, It could be observed a drop in body temperature from 38°C to 22°C, almost reaching the ambient temperature level, which oscillated between 28.5°C and 21°C, throughout the entire decomposition process (Fig. 1).

Very few flies were observed immediately following death, including the calliphorids *Lucilia eximia* and *Cochliomyia macellaria*, and *Bithoracochaeta* sp. (Muscidae). The initial egg masses were found in the mouth, 6 hours after death, and the eclosion of the eggs occurred 16 hours later. In the laboratory, it could be confirmed that these were *L. eximia* eggs. At the end of this stage, *L. eximia* larvae (L1 and L2) were found (Table III, IV). Epiponini wasps were found sucking the liquids drained from the wound.

Bloated (Day 1 through Day 5).

As the body temperature varied considerably during the days of heavy rain, ranging from 30.5 to 10°C, there was a corresponding fluctuation in the degree of bloating of the carcass (Fig. 1). When this peak was reached, the temperature began to drop until reaching 15°C, giving way the active

stage. The carcass weight was constant through the beginning of the bloated stage (Fig. 3).

In spite of the use of cages to protect the carcass, vultures pecked at the carcass in its bloated stage. As a result, the body experimented an early escape of gases. However, new attacks by vultures could be avoided, allowing the decomposition process and the colonization by insects to continue. Presence of larvae (L2 and L3) of Calliphorids *L. eximia*, *C. macellaria*, *Chrysomya albiceps*, *Hemilucilia semidiaphana*, *Chrysomya megacephala* and Silphid *Oxelytrum discicolle*, as well as their adults and of the beetle *Anotylus* sp. (Staphylinidae) in the mouth, eyes, nostrils and anus of the carcass is highlighted (Table II, III, IV).

Adults of other species not collected in the Fresh phase were: calliphorids *C. albiceps*, *C. macellaria*; muscids *Biopyrellia bipunctata*, *Musca domestica*, *Ophyra aenescens*, *Brontaea normata*, *Stomoxys calcitrans*; hymenopterans *Monomorium* sp. (Formicidae), *Melipona* sp. (Apidae) and hemipterans Coreidae (Table I). In this stage, ovoposition by flies under the body, on the back, head, ears and anus increased. Predators began to be observed, ants of the genus *Monomorium* sp. and *O. discicolle* adults, nesting under the body. Epiponini wasps and the staphilinids *Anotylus* sp. and *Paederomimus* sp. (Table I) feeding on the carcass, eggs and larvae.

Active Decay (Day 6 through Day 9).

The highest peak in temperature (29°C) occurred in the middle of the stage, during the colonization of larvae in the soft tissues of the entrails, with intermediate temperatures of 4°C with drops intermediate of up to 4°C, due to the strong rains (Fig. 1). The weight of the carcass dropped drastically, going from 22 Kg to 8 Kg during the consumption of the tissues by the larvae (Fig. 2).

The intense larval activity in the entrails, expected at the beginning of this stage, was not recorded until the middle of his duration since the attack by vultures accelerated the exposure of the entrails but not the colonization of the larvae in those tissues. The presence of 3rd instar larvae of the species *Comptosomyiops verena*, *Musca domestica* and L1, L2 and L3 of *Ophyra aenescens* (Muscidae) (Table III) is worth noting. Other species and their immature stages, *C. macellaria*, *C. albiceps*, *L. eximia* and *H. semidiaphana* were collected (Table IV).

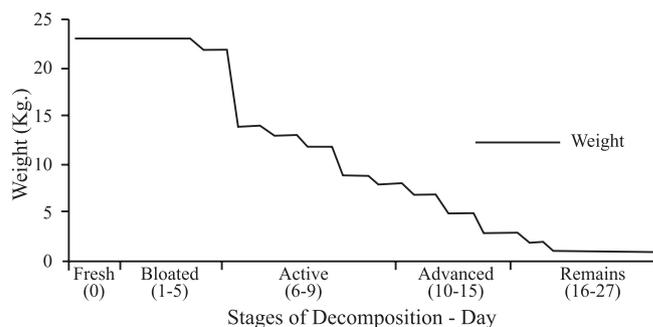


Fig. 2. Daily weight variations related with stage of decomposition.

Table I. Succession of insects associated with exposed carcasses in Colombia's Andean Coffee Region.

Order	Family	Species	Ecol.Cat.	Decomposition stages-Day									
				Fresh		Bloated		Active		Advanced		Remains	
				0	1-5	6-9	10-15	16-26					
Diptera	Muscidae	<i>Bithoracochaeta</i> sp.	S	X	X	X							
Hymenoptera	Vespidae	Epiponini	P,S	X	X	X							
Diptera	Calliphoridae	<i>Lucilia eximia</i> (Wiedemann, 1819)	N	X	X X	X X	X X	X					
		<i>Chrysomya megacephala</i> (Fabricius, 1794)	N			X							
		<i>Hemilucilia semidiaphana</i> (Rondani, 1850)	N		X	X	X	X					
Hemiptera	Coreidae	Unidentified	In		X								
Diptera	Muscidae	<i>Biopyrellia bipuncta</i> (Wiedemann, 1830)	S		X								
		<i>Brontaea normata</i> (Bigot, 1885)	S		X								
		<i>Musca domestica</i> Linnaeus, 1758	S		X	X	X X						
Coleoptera	Staphylinidae	<i>Anotylus</i> sp.	P		X	X	X						
		<i>Paederomimus</i> sp.	P		X	X	X						
Hymenoptera	Apidae	<i>Melipona</i> sp.	P,S		X	X	X						
	Formicidae	<i>Monomorium</i> sp.	P		X	X	X						
Diptera	Calliphoridae	<i>Chrysomya albiceps</i> (Wiedemann 1819)	N, P		X	X X	X X	X					
		<i>Cochliomyia macellaria</i> (Fabricius 1775)	N		X	X X	X X	X	X				
	Muscidae	<i>Stomoxys calcitrans</i> (Linnaeus, 1758)	S		X	X	X						
		<i>Ophyra aenescens</i> (Wiedemann 1830)	S		X	X	X	X	X	X			X
Coleoptera	Silphidae	<i>Oxelytrum discicolle</i> (Brullé, 1840)	N, P		X	X X	X	X	X				X
Diptera	Calliphoridae	<i>Paralucilia fulvinota</i> (Bigot, 1877)	N			X							
	Syrphidae	<i>Allograpta obliqua</i> Say 1823	In			X							
	Tephritidae	<i>Urophora</i> sp.	S			X							
	Otitidae	<i>Acrosticta</i> sp.	S			X		X					
Coleoptera	Histeridae	<i>Hister</i> sp.	P			X	X					X	
	Staphylinidae	<i>Philonthus</i> sp.	P			X	X						
Acari	Unidentified	Unidentified	In			X	X						
Diptera	Muscidae	<i>Haematobia irritans</i> (Linnaeus, 1758)	S			X						X	
Coleoptera	Dermestidae	<i>Dermestes</i> sp.	N			X	X					X	
Diptera	Piophilidae	<i>Stearibia nigriceps</i> (Meigen, 1826)	N,S			X	X						X
	Calliphoridae	<i>Compsomyiops verena</i> (Walker, 1849)	N				X	X					
Coleoptera	Chrysomelidae	Unidentified	In				X						
	Scarabaeidae	<i>Onthophagus</i> sp.	S				X						
	Staphylinidae	<i>Ambodina</i> sp.	P				X						
		<i>Styngetus</i> sp.	P				X						
Hemiptera	Mesoveliidae	Unidentified	In				X						
	Rhopalidae	Unidentified	In				X						
	Scutelleridae	Unidentified	In				X						
Diptera	Fanniidae	<i>Fannia</i> sp.	N						X				

N: Necrophagous, S: Saprophagous, P: Predator, In: Incidental, A: Adult, L: Larvae.

Adults that were not found in the bloated phase: Coleopterans *Dermestes* sp. (Dermestidae) and *Hister* sp. (Histeridae); dipterans *Paralucilia fulvinota* (Calliphoridae), *Stearibia nigriceps* (Piophilidae), *Acrosticta* sp. (Ulidiidae), *Allograpta obliqua* (Syrphidae), *Urophora* sp. (Tephritidae), muscids *Haematobia irritans*, *Stomoxys calcitrans*, and mites unidentified (Table I). Migration under the body of *L. eximia* and *H. semidiaphana* larvae was observed at the beginning of the stage. *C. albiceps* migrated at the end and *C. macellaria* larvae were found approximately 1m. from the body. Histerids (*Hister* sp.) were found buried under the body and dermestids (*Dermestes* sp.) were found under the very dry skin.

Advanced Decay (Day 10 through Day 15).

The days were hot and sunny during this stage, which the

body temperature fluctuated between 22°C and 37°C (Fig. 1). The weight went from 8 Kg to 3 Kg by the end of the stage, due to both the consumption of the remaining soft tissues by the larvae as well as to the dehydration of the remains (Fig. 2).

The number of Diptera larvae associated with the carcass decreased considerably and the diversity of the beetles increased (Table I, II). The presence of adult *Ambodina* sp. (Staphylinidae), as well as immature stages (L2, L3, prepupae, pupae) of *C. macellaria*, *O. aenescens*, *L. eximia*, *C. albiceps*, *H. semidiaphana*, *Fannia* sp. and *Compsomyiops verena* were worth noting (Table III, IV). The number of adult Diptera decreased, while the number of adult *O. aenescens*, which were found sucking liquids from the carcass and ovopositing under the body, increased.

Table II. Percentage of adult dipterans and coleopterans of forensic importance collected in exposed carcass in Colombia's Andean Coffee Region at each stage of decomposition.

Order	Family	Specie	Fresh (Day 0)	Bloated (Day 1-5)	Active (Day 6-9)	Advanced (Day 10-15)	Remains (Day 16-26)
Diptera	Calliphoridae	<i>Cochliomyia macellaria</i>	1,190	5,155	1,715	1,616	2,299
		<i>Chrysomya albiceps</i>	0,000	0,825	0,202	0,492	0,000
		<i>Lucilia eximia</i>	1,190	1,031	0,000	0,070	0,000
		<i>Paralucilia fulvinota</i>	0,000	0,000	0,101	0,000	0,000
	Muscidae	<i>Bithoracochaeta</i> sp.	1,190	0,206	0,101	0,000	0,000
		<i>Ophyra aenescens</i>	0,000	1,649	0,807	4,076	0,000
		<i>Musca domestica</i>	0,000	1,237	0,303	0,632	0,000
		<i>Stomoxys calcitrans</i>	0,000	0,206	0,202	0,351	4,598
		<i>Biopyrellia bipuncta</i>	0,000	0,206	0,000	0,000	0,000
		<i>Brontaea normata</i>	0,000	0,206	0,000	0,000	0,000
		<i>Haematobia irritans</i>	0,000	0,000	0,101	0,000	1,149
	Piophilidae	<i>Stearibia nigriceps</i>	0,000	0,000	0,303	0,422	0,000
	Syrphidae	<i>Chamaesyphus</i> sp.	0,000	0,000	0,303	0,000	0,000
	Otitidae	<i>Acrosticta</i> sp.	0,000	0,000	0,101	0,070	0,000
Tephritidae	<i>Urophora</i> sp.	0,000	0,000	0,101	0,000	0,000	
Coleoptera	Staphylinidae	<i>Anotylus</i> sp.	0,000	1,856	0,303	0,281	0,000
		<i>Philonthus</i> sp.	0,000	0,000	0,303	0,632	0,000
		<i>Paederomimus</i> sp.	0,000	0,000	0,202	0,000	0,000
		<i>Ambodina</i> sp.	0,000	0,000	0,000	0,422	0,000
		<i>Styngetus</i> sp.	0,000	0,000	0,000	0,202	0,000
	Silphidae	<i>Oxelytrum discicolle</i>	0,000	0,206	3,633	1,968	0,000
	Dermestidae	<i>Dermestes</i> sp.	0,000	0,000	0,404	7,238	8,046
	Histeridae	<i>Hister</i> sp.	0,000	0,000	0,404	1,405	0,000
	Scarabaeidae	<i>Onthophagus</i> sp.	0,000	0,000	0,000	0,141	0,000
	Chrysomelidae	Unidentified	0,000	0,000	0,000	0,070	0,000

Remains (Day 16 through Day 27).

The carcass did not undergo many changes in this stage. The body temperature oscillated 2 to 3 degrees higher than the ambient temperature (Fig.1). The weight went from 3 Kg to 1 Kg (Fig. 2).

The different species that were found in this stage are: *S. nigriceps* L2 and L3, *O. aenescens* L1, L2 and L3, *O. discicolle* larvae and *Dermestes* sp. and *Hister* sp. adults (Table IV).

Succession Matrix (Presence-Absence).

The succession matrix was created based on larvae of the following dipteran species: *L. eximia*, *H. semidiaphana*, *C. macellaria*, *C. megacephala*, *C. albiceps*, *C. verena*, *M. domestica*, *O. aenescens* and *S. nigriceps*; and adults of Coleoptera species *Anotylus* sp., *O. discicolle* (larvae and adult), *Paederomimus* sp., *Philonthus* sp., *Dermestes* sp., *Hister* sp. and *Ambodina* sp., which were the most representatives species of the succession and also were the colonizing species (Table IV).

DISCUSSION

Five stages of decay were observed: fresh, bloated, active, advanced, and remains. Their delimitation was done

considering the changes in the carcass's appearance, the body temperature and the presence of insects that marked these types of processes. Even so, the body temperature throughout the entire process oscillated almost with the ambient temperature. Also, the decomposition features for each of the stages were not clearly observed, which made it difficult to define the exact transition between the stages of decomposition. This was probably caused by the almost constant rain regime during the collections in October and November, the period of concentrated rain in the study area, suggesting that the duration of each stage can depend on the ambient conditions, which can affect the rate of decomposition (Schowalter 2000).

In tropical ecosystems, the rainfall pattern is more variable (Mavárez-Cardozo *et al.* 2005) and one of the factors that retard the decomposition is water, because body heat is lost twice as quickly as in the air (Smith 1986). Likewise, the insects associated with the corpse die by drowning affecting time of decomposition.

Davis & Goff (2000) concluded that water is a key factor in the differences between intertidal and terrestrial decomposition, demonstrating that it limits the access of carrion arthropods and reinforcing the hypothesis that the decomposition patterns and the organisms found in bodies vary according to the habitat, temperature and degree of exposure

Table III. Percentage of immature dipterans and coleopterans collected in exposed carcass in Colombia's Andean Coffee Region at each stage of decomposition.

Order	Family	Specie	Fresh (Day 0)	Bloated (Day 1-5)	Active (Day 6-9)	Advanced (Day 10-15)	Remains (Day 16-26)
Diptera	Calliphoridae	<i>Lucilia</i> sp. (Larvae I)	63,560	3,711	0,000	0,000	0,000
		<i>Lucilia eximia</i>	36,905	57,732	1,816	3,654	0,000
		<i>Cochliomyia macellaria</i>	0,000	15,463	68,416	35,067	0,000
		<i>Chrysomya albiceps</i>	0,000	6,186	9,788	1,195	0,000
		<i>Hemilucilia semidiaphana</i>	0,000	1,237	1,312	0,141	0,000
		<i>Chrysomya megacephala</i>	0,000	0,206	0,000	0,000	0,000
		<i>Compsomyiops verena</i>	0,000	0	0,404	0,070	0,000
	Muscidae	<i>Ophyra aenescens</i>	0,000	0,000	0,605	30,850	83,908
		<i>Musca domestica</i>	0,000	0,000	0,101	0,000	0,000
	Fanniidae	<i>Fannia</i> sp.	0,000	0,000	0,000	0,141	0,000
Coleoptera	Silphidae	<i>Oxelytrum discicolle</i>	0,000	0,206	0,000	0,141	0,000

to water. This fact may make these types of decomposition incomparable with decomposition occurring in other ambient conditions (e.g. dry season).

The fresh stage lasted one day, differing from most successional studies conducted in Colombia, where the duration was from 2 to 4 days (Martínez *et al.* 2007; Segura *et al.* 2009; Camacho 2005). Nonetheless, the duration of the other decomposition stages (bloating – 5 days, active – 4 days, advanced – 5 days and remains – 11 days) are within the range of days observed in those studies.

Lucilia eximia, *C. macellaria* and *Bithoracochaeta* sp. were the first species to colonize the carcass. However, *L. eximia* was the first to oviposit on the carcass, approximately 10 minutes after death, initially in the mouth and, subsequently, on the back and face. These results differ from those obtained in Medellín, where *Phaenicia sericata* was the first species to appear (Wolff *et al.* 2001; Pérez *et al.* 2005); *Calliphora nigribasis* and *Compsomyiops verena* in the Chingaza National Park in highlands ecosystem (Martínez *et al.* 2007); *Calliphora vicina* in Bogotá (Camacho 2005) and *C. macellaria* in Sucre (D. Grisales, unpublished data).

Of the species found here, some were already reported in Colombia associated with decaying bodies and for the same life zone of the study area, including *L. eximia* as a primary colonizer; *C. macellaria* as a secondary species, arriving at the bodies after these have been colonized by other species and *H. semidiaphana*, collected in carcasses exposed in the bloated through the advanced stage (Wolff & Vélez 2007), reports that are consistent with the behavior of these species in this study.

From the bloated through the advanced stage, *L. eximia*, *C. macellaria*, *C. albiceps*, and *H. semidiaphana*, in that order of importance, were mainly responsible for the removal of the carcass's tissues, with a great amount of *C. macellaria* found in the advanced stage, together with *Ophyra aenescens*. This muscid species was the dominant species in the remains stage, a finding not recorded in forensic entomology studies in Colombia, where the indicator species of this stage were

beetles Dermestids, Histerids, Staphylinidae and Silphidae (Wolff *et al.* 2001; Segura *et al.* 2009); Dipterans *Piophilina foveolata* (Piophilidae) (Pérez *et al.* 2005) and *Leptocera* sp. (Sphaeroceridae) (Martínez *et al.* 2007). This fact perhaps is due to the food preferences of this species, which feeds on the mucilaginous material (BOD) usually present in the advanced stage, but in this case it was more evident in the stage of remains.

In this study the carcass took 27 days to decompose. In the study conducted by Gomes *et al.* (2009), where the model and the study area were similar, the decomposition occurred in 28 days during the rainy season, while in studies conducted in the dry season, the total decomposition time was between 83 days (Martínez *et al.* 2007) and 45 days (Carvalho & Linhares 2001), depending on the altitude, which influences both the humidity and the diversity of the insects.

Among the forensically important beetles, *Anotylus* sp., *Paederomimus* sp. and *Philonthus* sp., (Staphylinidae) adults were collected in this study and were also reported by Nuorteva (1977) in Finland. The predators *Ambodina* sp. aff. (Staphylinidae), *Dermestes* sp. (Dermestidae), *Hister* sp. (Histeridae) and *O. discicolle* (Silphidae) arrived at the carcass once there were larvae and other insects to feed on. The most representatives were *Anotylus* sp. and *O. discicolle*, but only the latter, the most abundant in the process, was collected both as immatures and adults. This species is predatory and necrophagous, found feeding mainly under the carcass and in the skull. The feeding habits of adult and immature specimens of some species of necrophagous beetles differ, due to the competition with the Diptera that are more agile at colonizing a carcass quickly. Necrophagous beetles prefer to arrive at the carcass later, in drier stages, and the predator ones have the strategy of arriving before the fly larvae migrate far from the body (Olivera-Costa 2007). In a study conducted in the city of Curitiba (Brazil), where the fauna of Coleoptera was collected from a pig carcass (Mise *et al.* 2007), *O. discicolle* was found from the bloated phase until the end of the decomposition. This observation coincides

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